# Laboratory assays for the measurement of Factor VIII and FIX

#### Anna Pavlova

Institute of Experimental Haematology and Transfusion Medicine University Clinic Bonn

8<sup>th</sup> ECAT Participants' Meeting

8<sup>th</sup> and 9<sup>th</sup> of November 2012, Leiden, The Netherlands



clinical severity of haemophilia A/B

circulating activity of FVIII/IX

severe (FVIII/IX:C <1.0 IU/dI) moderate (FVIII/IX:C 1.0–5.0 IU/dI) mild (FVIII/IX:C >5.0 IU/dI)

#### FVIII/IX activity measurement

#### Diagnosis

accurate diagnosis of FVIII/IX deficiency

- assessment of clinical severity
- identification of patients with sub-haemophilia
- identification carriers of haemophilia

#### Treatment

- monitoring of on-demand and prophylactic therapy
- prerequisite for patient-tailored treatment strategy
- monitoring postinfusion pharmacokinetics of concentrates

#### Manufacturers

potency labelling of clotting factors concentrates

### Laboratory analyses of FVIII/IX deficiency



aPTT - simple, rapid and highly reproducible

provide only a little information

Second-line coagulation testing



#### Alternative coagulation tests

Global hemostasis assays

Thrombin generation test (TGT) Thromboelastography (TEG) aPTT waveform analysis

kinetics of clot formation

### Ideal Laboratory FVIII/IX activity measuring assays

High Specificity - to assess unequivocally FVIII/IX in the presence of components which may be expected to be present

High Accuracy -the closeness between an accepted reference value and the value found.

High Precision –close results between a series of measurements obtained from multiple sampling of the same homogeneous sample

Repeatability

Reproducibility

Detection Limit -lowest amount of FVIII/IX in a sample which can be detected but not necessarily quantitated as an exact value (<1 IU/dL)</p>

Quantitation Limit -is the lowest amount of analyte in a sample which can be quantitatively determined with suitable precision and accuracy

Calibration (traceability to International Standard)

Compliance with international guidelines



FVIII/IX content in plasma or treatment concentrates

evaluated by activity-based assays

one-stage activated partial thromboplastin time (APTT) based clotting assay

chromogenic assay

#### One-stage vs Chromogenic Assay

#### **One-stage assay**

Plasma based system with native concentrations of coagulation factors

Based on measuring the activated partial thromboplastin aPTT) - endpoint: clot formation

Wide heterogeneity of available methods (reagents, alysers, deficient plasmas)

High inter- and intra laboratory variability

#### **Chromogenic** assay

Artificial system employing highly diluted plasma with high thrombin concentrations

Two sequential reactions : 1<sup>st</sup> – FXa forming ; 2<sup>nd</sup> –measurement of FXa by chromogenic substrate

Independence of clotting-specific reagents, easy to automate, only several commercial kits available

Less inter- and intra laboratory variability

Disadvantage

inability to measure very low levels of clotting factor activity

#### Variables influencing assays

32–68% of all laboratory errors occur in the preanalytical phase Bonini et al 2002

Pre-analytical Variables

sample collection- phlebotomy, blood collection systems,

sample handling- transport, exposure to heat, overall time to delivery, haemolysis, icterus and lipaemia, storage

Analytical

✓ quality of the reagents

Reference plasma

Factor VIII/IX-deficient plasma -the absolute absence of FVIII/IX, excess of all factors except FVIII, presence of VWF

aPTT reagent

✓ alterations in the procedures used in different laboratories

Detection system employed

version experience and precision of the personnel

one-stage activated clotting assay chromogenic assay

generally a good correlation between factor levels and clinical phenotype

Dicrepancies between measured factor VIII/IX levels and clinical phenotype



heterogeneity in bleeding pattern among patients with severe hemophilia

➢effect of FVIII replacement therapy in the absence of FVIII activity response

haemophilia B gene therapy – therapeutic effect already at FIX level of <1 IU/dL</p>

no standardized assays to assess patient response to bypassing agents Misdiagnose haemophilia with one of the methods

#### ¥

Mild haemophilia A patients with specific mutations

Potency labeling of newly developed recombinant products

#### Heterogeneity in bleeding pattern among patients with severe hemophilia

10–15% of severe hemophiliacs (factor activity<1 IU/ dL) differ considerably in their bleeding tendency possibly due to:

E.Santagostino et al 2010;van den Berg M et al 2008

Individual

prothrombotic mutations -FV Leiden or prothrombin G20210A

inter-individual variance in the pharmacokinetics of FVIII

von Willebrand antigen level (ABO blood group -patients with blood group O, have a 30% lower von Willebrand antigen level)

Gene defects

Contradictory data, more studies are needed

Laboratory

variations in FVIII/IX activity below detection limits (FVIII/IX<1IU/dl)

Standard clotting and chromogenic assays

precise low levels of FVIII/IX:C (<0.2 IU/dl) more consistent with severe clinical phenotype

Shima et al 2008

Can this problem be solved?

Application of two standard curves covering the lower and higher ranges of clotting factor activity

Optimised chromogenic assay -method based on commercially available reagents that claims a detection limit of 0.15 IU /dL

Yatuv R et al 2006

Can this problem be solved?

Global hemostasis assays

Clot waveform analysis

Thrombin generation

Thromboelastography

### Clot waveform analysis



discriminate between different levels of FVIII:C in severe HA (FVIII:C <1.0 IU/dl by conventional assays)</li>

•Shima M, et al 2002, Matsumoto T, et al 2006

detect variability in coagulation activity in individual patients

Nair SC et al 2010

monitoring the haemostatic effect of FVIII concentrate during ITI therapy

Kasuda S, et al 2004

assessment of clotting function in acquired haemophilia A

Matsumoto T, et al 2012

### Thrombin generation test



Dynamic assay – real time measurement of thrombin generation in PPP and PRP

Can TGT discriminate FVIII/IX levels <1IU/dl?

little information is available on the use of the TGT for assessing very low levels of FVIII:C

good correlation between clinical severity and PeakTh in hemophilia A, the TF-TGT did not offer any advantages for the evaluation of clinical severity

Beltran-Miranda 2005

> a modified TF/ELG-TGT is sensitive to very low levels of FVIII:C (app.0.2 IU/dl)

Matsumoto et al 2009

#### Thrombin generation test

individually assess bleeding risk in hemophilia patients or to define the phenotypic heterogeneity seen in these patients

Dargaud Y et al 2005

> monitor bleeding risk and response to treatment in acquired hemophilia

Spiezia L et al 2009

show inter- and intra-patient variability in response to treatment

Kennet G et al 2010

corelates with bleeding risk and could predict dosing of bypassing agents for hemophilia patients with high-titer inhibitors

Chitlur M 2012 Dargaud Y et al 2010;

### Thromboelastography



TEG or ROTEM - continuous assessment of the viscoelastic properties of a forming clot

Correlation of thromboelastography with clinical phenotype

Considerable heterogeneity in the thromboelastography patterns amongst patients with verified factor VIII levels < IU/dl</p>

Ingerslev J,et al 2003

Less pathological clotting profiles are associated with a less severe bleeding phenotype

Chitlur M,et al 2008

> predict the clinical response to bypassing agents in patients with inhibitors

Sorensen B, Ingerslev J 2004, 2007

### Global hemostasis assays in determination of low FVIII/IX levels (limitations)

still more expensive

labor -intensive

require specific equipment and qualifications

require a rigorous process of pre-analytical and analytical standardization

the correlation between test results and FVIII concentrations are not always conclusive

High intrapatient and interpatient variability

a high coefficient of variation of external quality assurance samples

limit their implementation within the routine of most clinical laboratories.

Discrepancy between chromogenic and clotting assays VIII levels chromogenic and clotting assays



NO consensus as to which method most accurately represents the FVIII cofactor function in vivo and gives the clinically relevant FVIII:C levels.



#### FVIII:C<sub>1st</sub> > FVIII:C<sub>chr</sub>

| Mutations   | FVIII: Ag<br>(U/dI) | FVIII:C1st<br>(IU/dI) | FVIII:Cchr<br>(IU/dI) | Ratio FVIII:C<br>chr/1st |
|-------------|---------------------|-----------------------|-----------------------|--------------------------|
| Tyr114Cys   | 25                  | 7                     | 4                     | 0,6                      |
| Thr118lle   | 11                  | 2                     | 1                     | 0,6                      |
| Asp203Val   | 8                   | 2                     | 1                     | 0,6                      |
| Thr275lle   | 35                  | 4                     | 1                     | 0,2                      |
| Asn280lle   | 15                  | 6                     | 3                     | 0,5                      |
| Arg531His*  | 60(59-61)           | 26(20-31)             | 13(12-14)             | 0,5                      |
| Arg531Cys*  | 28(45-15)           | 13(11-14)             | 5(4-7)                | 0,4                      |
| Val663Ala   | 54                  | 28                    | 18                    | 0,6                      |
| Arg1749His* | 60                  | 10                    | 5                     | 0,5                      |
| Pro1825ser  | 41(38-44)           | 15(14-16)             | 7(6-8)                | 0,5                      |
| Arg1941GIn  | 23                  | 7                     | 4                     | 0,5                      |

#### FVIII:C<sub>chr</sub> > FVIII:C<sub>1st</sub>

| Mutations   | FVIII: Ag<br>(U/dI) | FVIII:C1st<br>(IU/dI) | FVIII:Cchr<br>(IU/dI) | Ratio FVIII:C<br>1st/1chr |
|-------------|---------------------|-----------------------|-----------------------|---------------------------|
| Lys166Thr   | 21                  | 9                     | 18                    | 0,5                       |
| The295Ala   | 18                  | 9 (6-22)              | 17 (10-41)            | 0,5                       |
| Leu412Phe   | 18                  | 4                     | 10                    | 0,4                       |
| Arg527Trp** | 90 (61-138)         | 12 (8-23)             | 31 (20-52)            | 0,4                       |
| Val678Leu   | 94                  | 39                    | 78                    | 0,5                       |
| Met702Leu   | 10                  | 2                     | 6                     | 0,3                       |
| Glu720Lys*  | 88 (62-133)         | 24 (16 -35)           | 94 (54-120)           | 0,2                       |
| Tyr1680Phe  | 16 (10-39)          | 4 (2-7)               | 9 (4-13)              | 0,5                       |
| Arg1689His* | 113                 | 16                    | 41                    | 0,3                       |
| Leu1756Phe  | 17                  | 9                     | 19                    | 0,5                       |
| Leu1756Val  | 10                  | 3                     | 9                     | 0,4                       |
| Arg1781Gly  | 10                  | 4                     | 7                     | 0,5                       |
| Ser2119Tyr  | 15 (10-24)          | 3(2-5)                | 7 (5 -9)              | 0,4                       |

### Proposed Mechanism FVIII:C<sub>1st</sub> > FVIII:C<sub>chr</sub> Discrepancy



### Proposed Mechanism FVIII:C<sub>1st</sub> < FVIII:C<sub>chr</sub> Discrepancy



#### Conclusions (I)

Although the difficulties in standardisation, plasma coagulation assays are still the golden standard for measuring FVIII/IX levels

The improvement of the routine management of severe hemophiliacs requires an understanding of the clinical significance of extremely low levels of FVIII and FIX activites

Application of global haemostatic tests for measuring FVIII/IX, especially in levels < 1IU/dl still have to be demonstrated</p>

Application of global haemostatic tests might help in accurate and efficient monitoring of hemostatic response to bypassing agents in tratments hemophiliac patients with inhibitors

Better knowledge of the exact mechanism of application of different tests in hemophilic patients may improve individually tailored treatment strategies.

### Conclusions (II)

The FVIII:C one-stage and chromogenic assay discrepancies showed systematic differences

- due to the specific missense mutation
- due to assay procedures itself

Mutations can be used as a model to dissect the nature and mechanisms by which the various FVIII:C assays showed systematically discrepant results

➤ The initial diagnoses of non-severe HA phenotypes should be based on results from both, the FVIII:C one-stage and FVIII:C chromogenic assays.

## Thank you for your attention !